

Amendments to the Specification:

Please replace the paragraph on page 1, lines 3-4 with the following rewritten paragraph:

-- This filing is a divisional of commonly assigned, then co-pending application 09/970,446, filed October 3, 2001, now US Patent Number 6,670,146, which patent application claims benefit of US provisional patent application No. 60/239,023, filed October 4, 2000, each of which is incorporated by reference. --

Please replace the paragraph beginning on line 17 of page 12, with the following rewritten paragraph:

-- Stimulation of naïve naive T cells (CD4+CD62L+) using Antigen Presenting Cells (APC) in the presence of VIT D3 and DEX gives rise to: about 60% IL-10 positive cells (producing at least 500 ng of IL-10 per 10⁶ cells); about 10% IL-10 and IL-4 double positive cells (producing about 50 ng of IL-4 per 10⁶ cells); less than about 1% IL-5 positive cells (producing less than about 30 pg of IL-5 per 10⁶ cells); and less than about 5% IFN- γ positive cells (producing less than about 40 ng of IFN- γ per 10⁶ cells). The cytokine profiles produced by the cells after restimulation are evaluated after 6 h by FACS analysis, while the quantities are accumulated by the respective cell population in 1 ml for 48 h. --

Please replace the paragraph beginning on line 27 of page 12, with the following rewritten paragraph:

-- Alternatively, stimulation of naïve naive T cells using Antigen Presenting Cells (APC) in the presence of VitD3 and Dex + anti-IL-4 + anti-IL-12 + anti-IFN- γ gives rise to a population of cells: about 25-35% IL-10 positive cells (producing at least about 100 ng of IL-10 per 10⁶ cells); less than about 1% IL-4 positive cells (producing about 30-1000 pg of IL-4 per 10⁶ cells); less than about 1% IL-5 positive cells (producing less than about 30 pg of IL-5 per 10⁶ cells); less than about 1% IFN- γ positive cells (producing less than about 30 pg of IFN- γ per 10⁶ cells); and less than about 5% IL-2 positive cells (producing about 30-350 pg IL-2 per 10⁶ cells). --

Please replace the paragraph beginning on line 1 of page 13, with the following rewritten paragraph:

-- If the stimulation of naïve naive T cells is performed in the presence of anti-CD3 + anti-CD28 with VitD3 and Dex, the resulting population of cells is: about 70-75% IL-10 positive cells; less than about 2% IL-4 positive cells; less than about 1% IL-5 positive cells; less than about 1% IFN- γ positive cells; and less than about 2% IL-2 positive cells. Corresponding quantities of cytokines should be detected according to the cell types described immediately above. --

Please replace the paragraph beginning on line 8 of page 13, with the following rewritten paragraph:

-- Stimulation of naïve naive T cells using anti-CD3 + anti-CD28 in the presence of VitD3 and Dex + anti-IL-4 + anti-IL-12 + anti-IFN- γ gives rise to a population of: about 60% IL-10 positive cells; less than about 1% IL-4 positive cells; less than about 1% IL-5 positive cells; less than about 1% IFN- γ positive cells; and about 10% IL-2 and IL-10 double positive cells. --

Please replace the paragraph beginning on line 13 of page 13, with the following rewritten paragraph:

-- This data on the development of IL-10 only producing cells using Vitamin D3 and Dexamethazone plus antigenic stimulation of CD4+ T cells has also been reproduced in human systems. In this case there is some dependency on IL-4. Stimulation of purified human naïve naive T cells from cord-blood (CD4+CD45RA+) using L cells (expressing CD32, CD86) as APC plus anti-CD3 in the presence of VitD3 and Dex gives rise after one week of stimulation to a majority of IL-10 producing T cells: VitD3 is used at 2×10^{-8} M (2×10^{-9} M to 2×10^{-7} M) while Dex is used at 10^{-7} M (10^{-8} M to 10^{-6} M). In addition, stimulation of purified naïve naive T cells enriched from human peripheral blood (CD4+CD45RA+) using anti-CD3/+/ anti-CD28, in the presence of VitD3 and Dex gives rise after one week of stimulation to a majority of IL-10 producing T cells. --

Please replace the paragraph beginning on line 30 of page 24, with the following rewritten paragraph:

-- Peripheral blood mononuclear cells (PBMC) were isolated from peripheral blood, freshly collected into sodium citrate, by centrifugation on a Lymphoprep (Nycomed, Birmingham, UK) density gradient. Donors were healthy at the time of the study. PBMC were positively selected for CD8+ or CD4+ T cells using antibody-coated magnetic beads and Detach-a-beads (Dynal (UK), Wirral, GB) according to the manufacturer's guidelines. Alternatively, PBMC were depleted of CD4+ or CD8+ T cells by negative selection using antibody-conjugated magnetic beads to give CD8+ APC or CD4+ APC populations, respectively. CD4+ T cells were further subdivided into naïve naive CD45RA+ and antigen-experienced CD45RO+ populations using specific antibodies (PharMingen) and negative selection with magnetic beads. APC consisted predominantly of CD4low, CD14+ monocytes (13-28%), although a minor population (1.3-5%) of CD4low, CD14- cells is likely to contain DC. Isolated populations were washed and resuspended at 1×10^6 /ml in RPMI 1640 (Life Sciences, Abingdon, GB) containing 10% heat-inactivated FCS (PAA Laboratories, Oxford, GB), 2 mM L-glutamine, and 50 μ g/ml gentamycin (both from Life Sciences). Cell purity was assessed by flow cytometry using a FACScan (Becton Dickinson, Abingdon, GB) and Lysis 2 software. Anti-CD4 (Leu-3a) and CD14 (Leu-M3) antibodies were purchased

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from Becton Dickinson; CD3 (UCHT1), CD8 (UCHT4), isotype-matched control IgG1 (MOPC21), and IgG2b (MOPC141) antibodies were from Sigma (Poole, GB). --